

06-05-00

A

Docket Number 100-8024C/C1/D1

FILING BY "EXPRESS MAIL" UNDER 37 CFR 1.10

EL402023172US
Express Mail Label NumberJune 2, 2000
Date of DepositAddress to: Assistant Commissioner for Patents
Box Patent Application
Washington, DC 20231

UTILITY PATENT APPLICATION TRANSMITTAL AND FEE SHEET

Transmitted herewith for filing under 37 CFR §1.53(b)(1) is a **divisional** of prior Application No. 09/072,278, filed May 4, 1998.

Applicant (or identifier): RICHARD SEDRANI AND VALERIE QUESNIAUX RYFFEL

Title: RAPAMYCIN ASSAY

Enclosed are:

1. ☒ Specification (Including Claims and Abstract) - 25 pages
 2. ☒ Drawings - 8 sheets
 3. ☐ Declaration and Power of Attorney
 - a. ☐ Newly executed (original or copy)
 - b. ☒ Copy from a prior application (signed or with indication that original was signed)
 - i. ☐ Deletion of Inventors
Signed statement attached deleting inventor(s) named in the prior application
 4. ☒ Incorporation By Reference
The entire disclosure of the prior application, from which a copy of the Declaration and Power of Attorney is supplied under Box 3b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.
 5. ☐ Microfiche Computer Program (appendix)
 6. ☐ Nucleotide and/or Amino Acid Sequence Submission
 - ☐ Computer Readable Copy
 - ☐ Paper Copy
 - ☐ Statement Verifying Identity of Above Copies
 7. ☐ Preliminary Amendment
 8. ☐ Assignment Papers (Cover Sheet & Document(s))
 9. ☐ English Translation of
 10. ☐ Information Disclosure Statement
 11. ☐ Certified Copy of Priority Document(s)
 12. ☒ Return Receipt Postcard
 13. ☐ Other:
- ☒ The right to elect an invention or species that is different from that elected in parent Application No. 09/072,278 in the event of a restriction or election of species requirement that is identical or substantially similar to that made in said parent application is hereby reserved.

Filing fee calculation:

jc831 U.S. PTO
06/02/00jc825 U.S. PTO
09/585743
06/02/00

09/072,278-060000

- ☐ Before calculating the filing fee, please enter the enclosed Preliminary Amendment.
☐ Before calculating the filing fee, please cancel claims

Basic Filing Fee								\$	690
Multiple Dependent Claim Fee (\$ 260)								\$	
Foreign Language Surcharge (\$ 130)								\$	
	For	Number Filed		Number Extra		Rate			
Extra Claims	Total Claims	14	-20	0	x	\$	18	=	\$
	Independent Claims	3	-3	0	x	\$	78	=	\$
TOTAL FILING FEE								\$	690

- ☒ Please charge Deposit Account No. 19-0134 in the name of Novartis Corporation in the amount of \$690. An additional copy of this paper is enclosed. The Commissioner is hereby authorized to charge any additional fees under 37 CFR §1.16 and §1.17 which may be required in connection with this application, or credit any overpayment, to Deposit Account No. 19-0134 in the name of Novartis Corporation.


Please address all correspondence to the address associated with Customer No. 001095, which is currently:

Thomas Hoxie
Novartis Corporation
Patent and Trademark Dept.
564 Morris Avenue
Summit, NJ 07901-1027

Please direct all telephone calls to the undersigned at the number given below and all telefaxes to (908) 522-6955.

Respectfully submitted,

Date: June 2, 2000

 June 2, 2000
Diane E. Furman
Attorney for Applicants
Reg. No. 31,104
Tel. No. (908) 522-6924

RAPAMYCIN ASSAY

This invention relates to monoclonal antibodies to rapamycin and rapamycin derivatives, which are useful, e.g., in assay kits for monitoring blood levels of drug.

Rapamycin is a macrolide antibiotic produced by Streptomyces hygroscopicus, which has been found to be pharmaceutically useful in a variety of applications, particularly as an immunosuppressant, e.g., for use in the treatment and prevention of organ transplant rejection and autoimmune diseases. Rapamycin, however, does exhibit side effects at higher dosages, and it has a somewhat variable bioavailability. Monitoring blood levels of rapamycin in patients being treated with rapamycin is thus very desirable in order to be able to regulate the dosage so as to maintain the minimum level sufficient for pharmacologic activity and to avoid any undue risk of side effects. The lack of a sensitive and reliable assay which can be performed quickly and easily in a clinical setting has been a major obstacle to the development of rapamycin as a pharmaceutical.

Previous efforts to develop assay kits for clinical monitoring of rapamycin have not been particularly successful. EP 041795, for example, describes a microbiological assay in which rapamycin concentration is measured as a function of antifungal activity. WO 92/02946 provides an assay system which measures rapamycin levels indirectly by measuring competition for binding to macrophilin. Both of these assays are cumbersome and not particularly sensitive. Even more importantly, both of these assays may have considerable variation under slightly different test conditions, making comparisons of test results from different hospitals difficult.

There have been no previous reports of monoclonal antibodies which recognize rapamycin. There are inherent difficulties in making monoclonal antibodies to rapamycin because rapamycin is not immunogenic and is itself extremely immunosuppressive.

Moreover, as the metabolites of rapamycin have not been well characterized in the literature, it is difficult to identify a monoclonal antibody capable of differentiating between rapamycin and its metabolites.

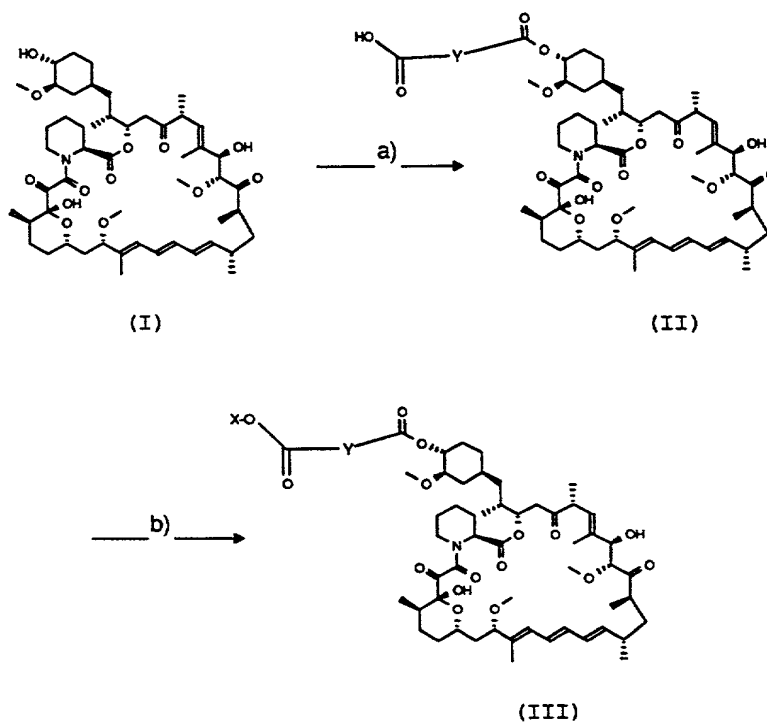
The present invention provides monoclonal antibodies which are highly sensitive to rapamycin. The antibodies of the invention are produced in response to inoculation with a novel immunogenic conjugate comprising a novel derivative of rapamycin linked to an immunogenic protein. Assay kits using these antibodies are well suited for use in a clinical setting and provide far more accurate and reproducible results than was previously possible. The antibodies are also useful in the purification and isolation of rapamycin.

Providing assay systems for immunosuppressive derivatives of rapamycin present similar challenges. Of particular interest are 40-O-derivatives of rapamycin, i.e., rapamycins which are O-substituted at the hydroxy on the cyclohexyl ring (position 40), e.g., as described in US 5 258 389 and PCT/EP 93/02604 (O-aryl and O-alkyl rapamycins) (both incorporated herein by reference); especially 40-O-alkylated rapamycins where the 40-O-substituent is alkyl or substituted alkyl; e.g., hydroxyalkyl, hydroxyalkoxyalkyl, acylaminoalkyl, or aminoalkyl, wherein "alk-" or "alkyl" refers to C₁₋₆ alkyl, branched or linear, preferably C₁₋₃ alkyl, in which the carbon chain may be optionally interrupted by an ether (-O-) linkage; most especially 40-O-(2-hydroxyethyl)-rapamycin, 40-O-(3-hydroxypropyl)-rapamycin, 40-O-[2-(2-hydroxy)ethoxy]ethyl-rapamycin, and 40-O-(2-acetaminoethyl)-rapamycin). Thus a further object of the invention is to provide monoclonal antibodies to such 40-O-derivatives. Such antibodies are useful in diagnostic assays and also in the purification and production of the derivatives.

The novel activated derivatives of rapamycin used to make the novel immunogenic conjugates of the invention are rapamycins which are linked through one of the hydroxy groups on the rapamycin, preferably the hydroxy group located on the cyclohexyl portion of the rapamycin (position 40) or the hydroxy at position 28, to an activated coupling group, i.e., a group capable of direct reaction with a protein to form a covalent linkage without the requirement for the use of a coupling agent (e.g., carbodiimide reagents) to

enable, effect, or promote the reaction with the protein. Preferably, the activated coupling group has an activated ester or carboxy group, i.e., of formula -CO-O-X where X is a carboxy activating group such as o- or p-nitrophenyl, 1-benztriazole, pentafluorophenyl, or (especially) N-succinimido. Other suitable activated coupling groups are, for example, i) activated dithio groups, e.g., of formula -S-S-Z wherein Z is a dithio activating group such as 2-pyridyl, which may be linked to the rapamycin; or ii) epoxy groups, e.g., epoxy methyl. The activated coupling group may be linked to the rapamycin by means of an ester, ether, amide, thio or other suitable linkage, but ester linkage is preferred. Most preferably, the activated coupling group contains a bis-ester moiety, e.g., succinyl, having an ester linkage to the rapamycin at one end and the activated ester or activated carboxy group at the other.

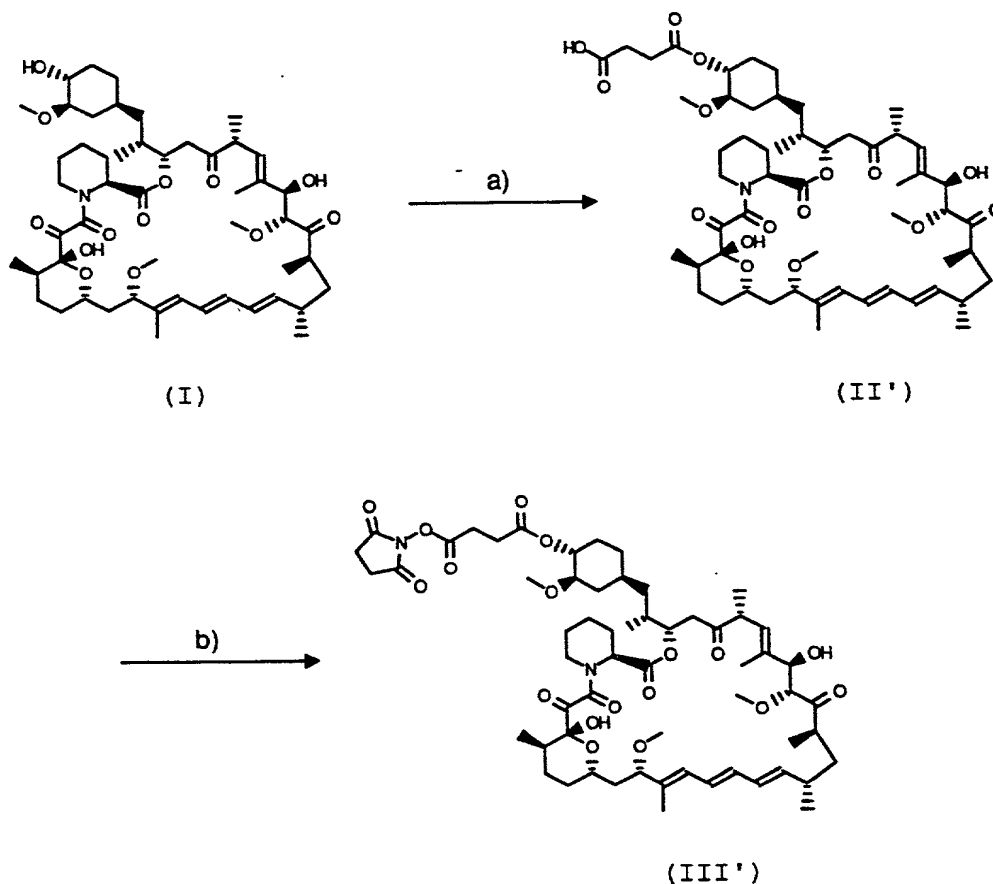
The preferred rapamycin derivatives of the invention are those of formula III below which are produced according to Reaction I:



Reaction I

wherein formula I is rapamycin, which is a) reacted with an acylating agent, e.g., a cyclic anhydride or a dicarboxylic acid (optionally in hemi-O-protected form), under suitable conditions and deprotection if necessary to yield the rapamycin of formula II, wherein Y is a spacer moiety, preferably a lower alkylene, e.g., C₂₋₆ alkylene, most preferably ethylene. This rapamycin of formula II is then b) activated by reaction with a carboxy activating group, e.g. of formula HO-X where X is as defined above, to yield the activated rapamycin of formula III.

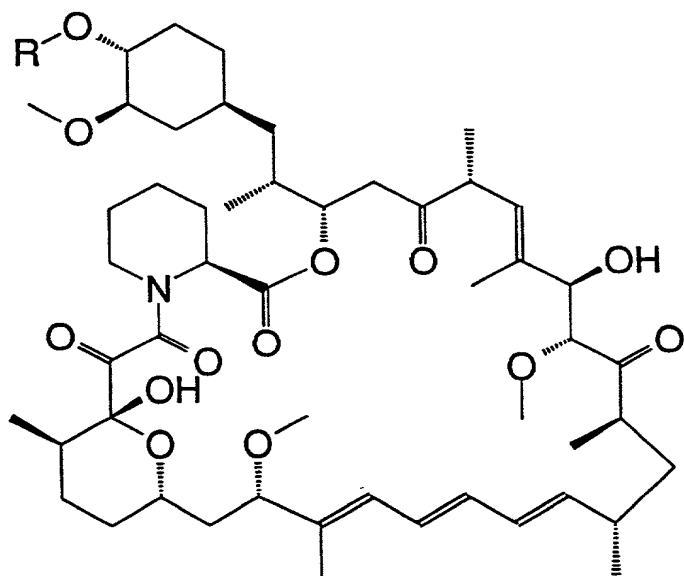
A preferred activated derivative of rapamycin is the succinimido derivative of formula III below, prepared, e.g., according to Reaction II:



Reaction II

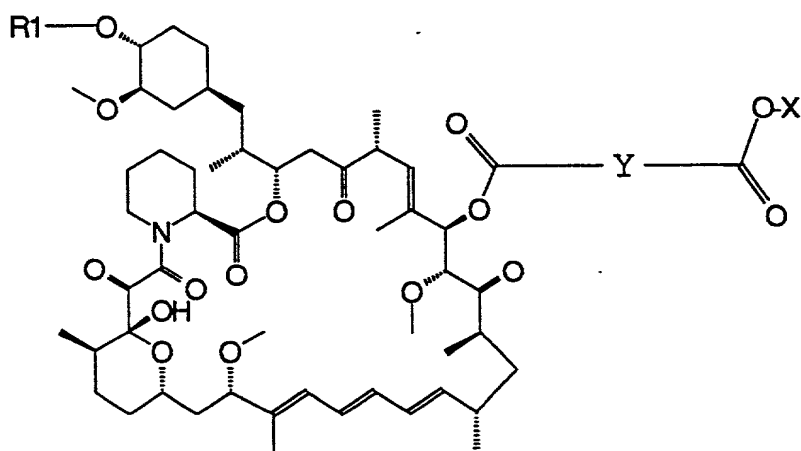
wherein formula I is rapamycin, which is a) O-acylated using succinic anhydride in the presence of DMAP and pyridine to form the rapamycin hemisuccinate of formula II' (40-O-(3-Carboxy)propanoyl-rapamycin); which is then b) activated with N-hydroxy succinimide in the presence of EDC, Et₃N, and CH₂Cl₂ to form the 40-O-succinimidooxysuccinyl rapamycin of formula III', e.g., as described more fully in example 1 below. Monoclonal antibodies produced using a hapten such as this which is linked through the 40-position will ordinarily be cross reactive between rapamycin and a 40-O-derivative of rapamycin, such as described above. Such monoclonal antibodies can be

It is in some cases desirable to have monoclonal antibodies capable of fine sensitivity to modifications in the cyclohexyl region, e.g., for distinguishing between rapamycin and the 40-O rapamycin derivatives, or for identifying metabolites in the cyclohexyl region. In such a case, the hapten is preferably linked through the 28-O position rather than the 40-O position. For example, the rapamycin derivative of formula A:



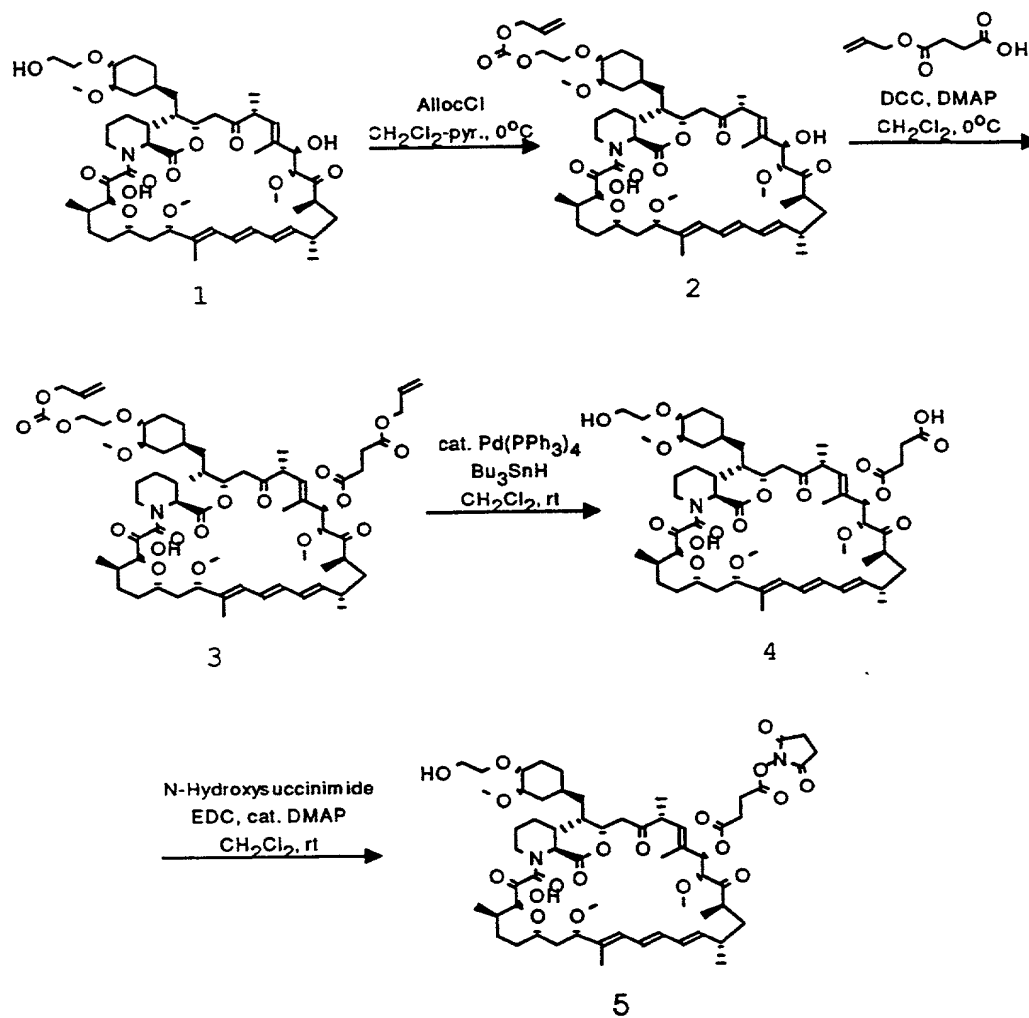
Formula A

wherein R is an O-protecting group, or a substituent as described above, e.g., hydroxyalkyl, hydroxyalkoxyalkyl, acylaminoalkyl, or aminoalkyl, optionally in protected form, is reacted according to Reaction I, deprotecting if necessary, to give the analogous 28-O activated hapten, for example a compound of formula B:



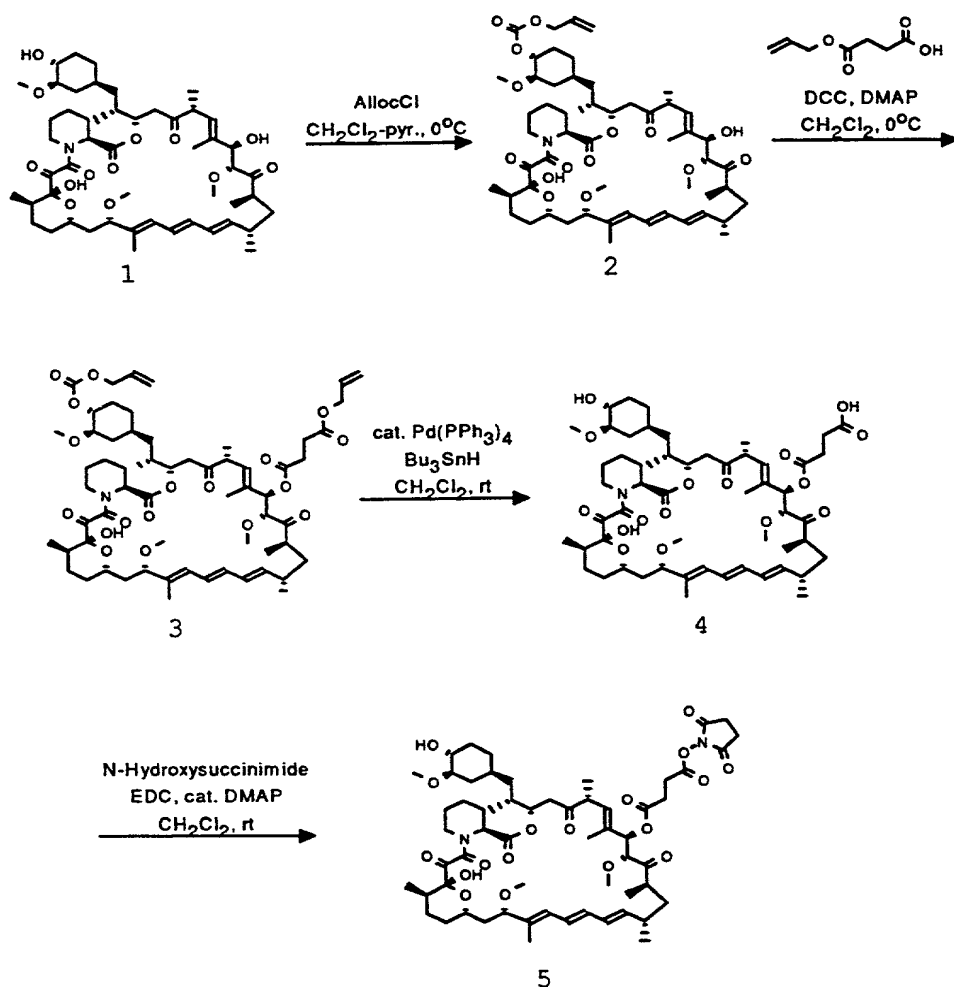
Formula B

wherein R1 is H, or an O-substituent as described above, e.g., hydroxyalkyl, hydroxyalkoxyalkyl, acylaminoalkyl, or aminoalkyl, Y is a linker moiety as defined above, and X is a carboxy activating group as defined above. In preparing this hapten, where R is an O-protecting group or an O-protected substituent, the acylating agent may optionally be, e.g., a dicarboxylic acid in hemi-O-protected form, so that following acylation, both O-protecting groups may be removed in one step prior to adding the carboxy activating group. For example, haptens for generating monoclonal antibodies capable of recognizing 40-O-(2-hydroxyethyl)-rapamycin, can be produced by protecting the primary hydroxy, acylating the hydroxy at position 28 with a dicarboxylic acid in hemi-O-protected form, deprotecting, and activating the carboxy group, e.g., according to Reaction III:



Reaction III

Similarly, rapamycin itself may be activated at the 28-O rather than the 40-O, by O-protecting the C40 hydroxy, acylating the hydroxy group in position 28 with a hemi-O-protected dicarboxylic acid, deprotecting, and activating the carboxy group, e.g., according reaction IV:



Reaction IV

The activated rapamycin or rapamycin derivative is then linked to a suitable immunogenic protein, e.g., bovine serum albumin (BSA), ovalbumin (OVA), or keyhole limpet hemocyanine (KLH) to form an immunogenic conjugate. Monoclonal antibodies are prepared using conventional methods, e.g., administering the novel immunogenic conjugate to a suitable animal species to effect immunogenic challenge and recovering antibody-producing cells sensitized to said conjugate; immortalizing said antibody producing cells by fusion with a suitable myeloma; and recovering the monoclonal antibody from a selected immortalized cell line thus established.

The antibodies of the invention may then be used in a suitable assay. Several possibilities would be clear to one skilled in the art. One approach is a competitive assay using antibody and a rapamycin tracer, for example wherein microtiter plates are coated with antibody and exposed to a competitor which is a labeled (e.g., fluoro- or radio-labeled, especially biotinylated) rapamycin, in the presence and absence of test fluid believed to possibly contain a rapamycin, e.g., plasma or whole blood from the patient. The plates are rinsed, and the amount of labeled competitor which has bound to the antibody is measured, which amount varies inversely with the amount of rapamycin in the test fluid. Another approach is an ELISA using antibody, a rapamycin protein conjugate, and a labeled (e.g., enzyme-labeled) tracer antibody recognizing murine IgG, for example wherein microtiter plates are coated with a rapamycin-protein conjugate (e.g., the immunogenic conjugate described above comprising a protein linked to rapamycin or a 40-O-alkylated rapamycin), exposed to antibody in the presence and absence of test fluid, rinsed, and antibody binding to the rapamycin conjugate detected by binding of the tracer antibody to the antibody bound to the rapamycin conjugate. Again, the amount of bound antibody will vary inversely with the amount of rapamycin in the test sample. In either case, the assay is standardized with test solutions containing known concentrations of rapamycin. An assay kit comprising (i) the monoclonal antibody of the invention, preferably in lyophilized form or coated onto a microtiter plate, and (ii) optionally also comprising either a rapamycin protein conjugate, optionally coated onto a plate, and/or a labeled rapamycin derivative, and (iii) further optionally comprising a rapamycin solution for standardization and instructions for use, is therefore provided. Such a kit is capable of

detecting rapamycin at concentrations of below 10 ng/ml, e.g., below 1 ng/ml, e.g., as low as 0.25-0.5 ng/ml.

The antibodies of the invention may be further characterized by their relative binding affinity to an immunosuppressive ascomycin, e.g., FK-506. FK-506 is an immunosuppressive macrolide having some structural similarity to rapamycin in the binding domain. Rapamycins (e.g., rapamycin and its immunosuppressant derivatives) and FK-506 both bind to macrophilins (FKBPs), and for both it is believed that macrophilin binding is a necessary but not a sufficient criteria for immunosuppressive activity. The effector region of rapamycin, however, is quite different from FK-506, and indeed, the two compounds have quite different mechanisms of activity. (FK-506 for example appears to cause immunosuppression primarily by suppressing IL-2 transcription, whereas rapamycin has no significant effect on IL-2 transcription.) Rapamycins can thus be characterized as having an FKBP binding domain and an effector domain, and a distinction can be made between rapamycin metabolites which are modified in the FKBP binding domain from those modified in the effector domain. This distinction can be made with the monoclonal antibodies of the invention by measuring the relative cross-reactivity of the monoclonal antibodies of the invention with FK-506 (cross-reactivity being measured, e.g., in a competitive ELISA): monoclonal antibodies having a high degree of cross reactivity (e.g., greater than 50%) recognize epitopes in the FKBP binding domain of rapamycin which is similar to FK-506; monoclonal antibodies with a low degree of cross reactivity (e.g., less than 20%, optimally less than 10%) recognize epitopes in the effector region, which is unique to rapamycins.

Antibodies of the invention can also be screened and characterized according to their ability to distinguish between rapamycin and a 40-O-derivative of rapamycin, e.g., as defined above. Where it is desired that the antibodies do not distinguish between rapamycin and a 40-O-derivative of rapamycin, antibodies are selected which show at least 70%, preferably greater than 90%, cross-reactivity between rapamycin and a 40-O-derivative thereof. In such a case, the hapten used to make the monoclonal antibody is preferably a 40-O-activated rapamycin, e.g., of formula III in Reaction I. Where it is

desired to distinguish between rapamycin and a 40-O-derivative or metabolite of rapamycin, antibodies are selected having less than 30%, preferably less than 10%, cross-reactivity thereto. In this case, the hapten used to make the antibody is preferably a 28-O-activated rapamycin or rapamycin derivative, e.g., of formula B.

EXAMPLE 1- Production of 40-O-activated rapamycin

a) *Production of 40-O-hemisuccinate of rapamycin*

To a stirred solution of 1.5 g (1.64 mmol) of rapamycin and 0.577 g (5.77 mmol) of succinic anhydride in 12 mL of pyridine is added 195 mg (1.64 mmol) of DMAP. The resulting mixture is stirred at ambient temperature for 19 h and concentrated under reduced pressure. The residue is dissolved in ethyl acetate and washed three times with water. The organic solution is dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure. The residue is purified by column chromatography on silica gel using 9:1 CH₂Cl₂-MeOH. The fractions containing the expected product are combined and purified once more by column chromatography on silica gel using 19:1 CH₂Cl₂-MeOH to afford, after removal of the solvent under reduced pressure, 40-O-(3-carboxy)propanoyl-rapamycin (the rapamycin hemisuccinate of formula II' supra) as a white foam showing the following characteristic spectroscopic properties:

¹H NMR (CDCl₃) δ 2.68 (7H, m, H33, H25 and O₂CCH₂CH₂CO₂H), 3.14 (3H, s and m, OCH₃ and H39), 3.34 (3H, s, OCH₃), 3.38 (3H, s, OCH₃), 4.68 (1H, m, H40), 4.72 (1H, broad s, 10-OH); MS (FAB) m/z 1036 ([M+Na]⁺), 982 ([M-CH₃O]⁺), 964 ([M-(CH₃O+H₂O)]⁺), 946 ([M-(CH₃O+2H₂O)]⁺).

b) *Production of 40-O-succinimidooxysuccinyl-rapamycin*

To a stirred solution of 120 mg (0.118 mmol) of the rapamycin hemisuccinate of step a), 16.5 μL (0.118 mmol) of Et₃N and 22.7 mg (0.118 mmol) of EDC in 8 mL of CH₂Cl₂ is added 13.6 mg (0.118 mmol) of N-hydroxysuccinimide. The resulting mixture is stirred for 18 h at room temperature, then diluted with ethyl acetate and washed twice with water. The organic solution is dried over anhydrous sodium sulfate, filtered and

concentrated under reduced pressure. The residue is purified by column chromatography on silica gel (ethyl acetate) to afford 40-O-succinimidooxysuccinyl-rapamycin (i.e., the compound of formula III' in Reaction II supra) as a white foam having the following characteristic spectroscopic properties:

^1H NMR (DMSO) δ 2.67 (2H, t, $\text{O}_2\text{CCH}_2\text{CH}_2\text{CO}_2$), 2.81 (7H, s, CH_3O and succinimide CH_2), 2.92 (2H, t, $\text{O}_2\text{CCH}_2\text{CH}_2\text{CO}_2$), 4.55 (1H, m, H40), 5.26 (1H, d, 28-OH), 6.43 (1H, s, 10-OH); MS (FAB) m/z 1133 ($[\text{M}+\text{Na}]^+$), 1111 ($[\text{M}+\text{H}]^+$), 1092 ($[\text{M}-\text{H}_2\text{O}]^+$), 1079 ($[\text{M}-\text{CH}_3\text{O}]^+$), 1061 ($[\text{M}-(\text{CH}_3\text{O}+\text{H}_2\text{O})]^+$), 1043 ($[\text{M}-(\text{CH}_3\text{O}+2\text{H}_2\text{O})]^+$).

EXAMPLE 2 - Production of 28-O-activated 40-O derivative of rapamycin

a) *28-O-hemisuccinate of 40-O-(2-hydroxyethyl)-rapamycin*

To a stirred, cooled (0°C) solution of 958 mg (1.00 mmol) of 40-O-(2-hydroxyethyl)-rapamycin in 2.2 mL of 10:1 methylene chloride-pyridine is added 0.160 mL (1.50 mmol) of allyl chloroformate. Stirring is continued at 0°C and two portions each of 0.080 mL (1.00 mmol) of pyridine and 0.053 mL (0.50 mmol) of allyl chloroformate are added after 3 hours and 4 hours respectively. After the last addition of reagents, stirring was continued for one more hour, and the reaction is quenched with 1M aqueous sodium bicarbonate. The resulting mixture is extracted three times with ethyl acetate. The organic solution is washed successively with 1N aqueous hydrochloric acid, 1N aqueous sodium bicarbonate and saturated brine, then dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure. The residue is purified by column chromatography on silica gel (50:50 hexane-ethyl acetate) to afford the allyloxycarbonyl protected compound (formula 2 in reaction III above) as a white foam.

To a stirred, cooled (0°C) solution of 208 mg (0.200 mmol) of this product in 2 mL of methylene chloride is added 2.4 mg (0.020 mmol) of DMAP and 82 mg (0.400 mmol) of DCC, followed by a solution of 63 mg (0.400 mmol) of monoallylsuccinate in 0.5 mL of methylene chloride. The reaction mixture is stirred at 0°C for 14 hours and the resulting suspension was filtered through a fritted glass funnel. The organic solution is concentrated

under reduced pressure, and the residue is purified by column chromatography on silica gel (30:70 hexane-ethyl acetate) giving the product (formula 3 of reaction III above) as a white foam.

To a stirred solution of 177 mg (0.150 mmol) of this product in 5 mL of methylene chloride is added 17.3 mg (0.015 mmol) of tetrakis(triphenylphosphine)palladium and 0.080 mL (0.3 mmol) of tributyltin hydride. The yellow solution is stirred for 2 hours at ambient temperature and diluted with ethyl acetate, washed once with cold 2N aqueous citric acid and three times with saturated brine, dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure. Column chromatography on silica gel (85:15 ethyl acetate-methanol) affords the hemisuccinate (formula 4 of reaction III) as a pale yellow oil.

b) *28-O-succinimidooxysuccinyl-40-O-(2-hydroxyethyl)-rapamycin*

A solution of 53 mg (0.050 mmol) of the hemisuccinate of step a) in 2.5 mL of methylene chloride is treated with 2 mg of DMAP, 24 mg (0.125 mmol) of EDC and 14 mg (0.125 mmol) of N-hydroxysuccinimide. After stirring for 2 hours at ambient temperature, the reaction is quenched with 1N aqueous sodium bicarbonate. The mixture is extracted with three portions of ethyl acetate. The organic solution is washed with aqueous sodium bicarbonate and brine, dried over anhydrous sodium bicarbonate, filtered and concentrated to give the title activated hapten (the compound of formula 5 of reaction III), which is used for the preparation of the protein-hapten conjugate without further purification, and which has the following characteristic spectroscopic properties:

^1H NMR (CDCl_3) δ 2.43 (1H, dd, H33a), 2.50-2.98 (10H, m, H25, H33b, succinate hydrogens, succinimide hydrogens), 3.58 (2H, m, H6b, 1 hydroxyethyl H), 3.68 (3H, m, H16, 2 hydroxyethyl H), 3.81 (2H, m, H14, 1 hydroxyethyl H), 3.93 (1H, d, H27), 5.28 (2H, m, H2, H30), 5.34 (1H, d, H28); MS (FAB) 1161 ($[\text{M}+\text{Li}]^+$).

EXAMPLE 3- Production of 28-O-activated rapamycin

a) *28-O hemisuccinate of rapamycin*

To a stirred, cooled (0°C) solution of 914 mg (1.00 mmol) of rapamycin in 2.2 mL of 10:1 methylene chloride-pyridine is added 0.212 mL (2.00 mmol) of allyl chloroformate. After 3 hours 0.080 mL (1.000 mmol) of pyridine and 0.053 mL (0.50 mmol) of allyl chloroformate are added. Stirring is continued for one more hour, and the reaction is quenched with 1M aqueous sodium bicarbonate. The resulting mixture is extracted three times with methyl-t-butylether. The organic solution is washed successively with cold 1N aqueous hydrochloric acid, 1N aqueous sodium bicarbonate and saturated brine, then dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure. The residue is purified by column chromatography on silica gel (70:30 hexane-ethyl acetate) to afford the allyloxycarbonyl protected compound (formula 2 in reaction IV) as a white foam.

To a stirred, cooled (0°C) solution of 400 mg (0.400 mmol) of this product in 5 mL of methylene chloride is added 4.8 mg (0.040 mmol) of DMAP and 164 mg (0.800 mmol) of DCC, followed by a solution of 127 mg (0.800 mmol) of monoallylsuccinate in 1 mL of methylene chloride. The reaction mixture is stirred at -15°C for 14 hours and the resulting suspension is filtered through a fritted glass funnel. The organic solution is concentrated under reduced pressure and the residue is purified by column chromatography on silica gel (40:60 hexane-methyl-t-butylether) giving the compound of formula 3 of Reaction IV as a white foam.

To a stirred solution of 285 mg (0.250 mmol) of this product in 5 mL of methylene chloride is added 28.8 mg (0.025 mmol) of tetrakis(triphenylphosphine)palladium and 0.133 mL (0.5 mmol) of tributyltin hydride. The yellow solution is stirred for 1 hour at ambient temperature and diluted with methyl-t-butylether, washed twice with cold 2N aqueous citric acid and three times with saturated brine, dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure. Column chromatography on silica gel (90:10-60:40 methyl-t-butylether-methanol) affords the 28-O rapamycin hemisuccinate (the compound of formula 4, reaction IV) as a pale yellow oil.

b) 28-O-succinimidooxysuccinyl-rapamycin

A solution of 51 mg (0.050 mmol) of the product of step a) in 2 mL of methylene chloride is treated with 2 mg of DMAP, 24 mg (0.125 mmol) of EDC and 14 mg (0.125 mmol) of N-hydroxysuccinimide. After stirring for 4 hours at ambient temperature, the reaction is quenched with 1N aqueous sodium bicarbonate. The mixture is extracted with three portions of methyl-t-butylether. The organic solution is washed with aqueous sodium bicarbonate and brine, dried over anhydrous sodium bicarbonate, filtered and concentrated to give the activated title compound, which is used for the preparation of the protein-hapten conjugate without further purification and which exhibits the following characteristic spectroscopic properties:

^1H NMR (CDCl_3) δ 2.43 (1H, dd, H33a), 2.55-3.02 (11H, m, H25, H33b, H39, succinate hydrogens, succinimide hydrogens), 3.56 (1H, m, H6b), 3.68 (1H, dd, H16), 3.83 (1H, m, H14), 3.93 (1H, d, H27), 5.28 (2H, m, H2, H30), 5.34 (1H, d, H28); MS (FAB) 1117 ($[\text{M}+\text{Li}]^+$).

EXAMPLE 4 - Production of immunogenic conjugates

a) *40-O-linked rapamycin conjugates*

17.4 mg of the 40-O-activated rapamycin of example 1 is dissolved in 400 μl DMF or DMSO. 120 μl (i.e., containing 5.22 mg activated rapamycin) of this solution is added dropwise with vigorous stirring to a solution containing 8 mg of KLH in 2 ml 0.1 M NaHCO_3 buffer (pH 7.7). The reaction mixture is stirred for 2 hours at room temperature, and the resulting rapamycin-KLH conjugate is purified by dialysis at 4°C against 5 l PBS, 3X over 48 hours. The conjugate is optionally further concentrated by centrifugation using microconcentrator tubes. Rapamycin-BSA and rapamycin-OVA conjugates are prepared in the same manner, substituting BSA or OVA respectively for KLH in the above procedure.

b) *28-O-linked (optionally 40-O-alkylated) rapamycin conjugates*

5 mg of the 28-O-activated compound of example 2 is dissolved in 2ml DMSO and added dropwise with vigorous stirring to a solution containing 5mg KLH in 1ml 50mM phosphate buffer (PH 7.3). The reaction mixture is stirred for 2 hours at room

temperature, and the resulting conjugate is purified by dialysis at 4°C against 21 PBS, 3X over 48 hours. Conjugates with BSA and OVA conjugates are prepared in the same manner. Rapamycin is conjugated to KLH, BSA, and OVA through position 28 using the 28-O activated compound of example 3, following the same procedure.

EXAMPLE 5 - Production of monoclonal antibody

a) *Monoclonal antibody to rapamycin*

Monoclonal antibody is produced by using conventional techniques, essentially as described by Köhler and Milstein in Nature 256: 49. Female Balb/C mice (20-25 g) each receive 10 or 50 µg of the 40-O-linked rapamycin-KLH immunogenic conjugate of example 4a) in 0.2 ml complete Freund adjuvant, administered by subcutaneous injection at four sites. After 2 weeks a second booster injection comprising the same amount of the immunogenic conjugate emulsified in 0.2 ml of incomplete Freund adjuvant is administered, again by s.c. injection. The presence of antibodies reactive to the antigen in the animals' blood serum is confirmed by direct ELISA as described in example 6 below. The mice may optionally be further selected for antibody to the effector region (low cross reactivity with FK-506) and to the FKBP binding region (high cross reactivity to FK-506). Figure 1, for example, shows titer curves for a mouse (M1) having a high level of antibody to the binding domain of rapamycin, and another mouse (M7) having relatively high levels of antibody to the effector domain. Mice displaying maximal blood serum levels of antibody of appropriate specificity receive booster injections comprising 10µg of antigen half i.p. half i.v. on day -3, and i.p. on day -2 and -1. On day 0. the mice are sacrificed and their spleen cells are isolated and fused with PAI-0 cells or other suitable myeloma line. The resulting hybridomae are cultured and selected using ELISA for expression of antibody having a high affinity to rapamycin.

b) *Monoclonal antibody to 40-(hydroxyethyl)-rapamycin*

Female Balb/C mice receive 10 or 50µg of the 40-(hydroxyethyl)-rapamycin KLH immunogenic conjugate of example 4b) in 0.2ml complete Freund's adjuvant by s.c.

injections on 4 points. After 2 weeks, a second injection (booster) comprising the same quantity of immunogenic conjugate emulsified in 0.2ml incomplete Freund's adjuvant is administered, again by s.c. injection. The presence of antibodies reactive to the antigen in the animal's blood serum is tested by direct ELISA as described below. In addition, mice are further selected for antibody to the region of the rapamycin molecule modified in the 40-O region by binding to a conjugate of BSA-rapamycin compared to a conjugate of BSA-40-O(2-hydroxyethyl)-rapamycin. Both antibodies which bind to the 40-O(2-hydroxyethyl)-rapamycin conjugate but do not bind to conjugated rapamycin, and antibodies which bind both to 40-O(2-hydroxyethyl)-rapamycin and rapamycin conjugates are obtained. Mice displaying maximal blood serum levels of antibody of appropriate specificity receive booster injections comprising 10µg of antigen half i.p. half i.v. on day -3, and i.p. on day -2 and -1. On day 0. the mice are sacrificed and their spleen cells are isolated and fused with PAI-0 cells. The resulting hybridoma are cultured and selected using ELISA for expression of antibody having a high affinity to 40-O-(2-hydroxyethyl)-rapamycin.

EXAMPLE 6 - Enzyme linked immunosorbent assay (ELISA)

a) *ELISA for rapamycin*

Microtiter plates are coated with 1-2 µg/ml rapamycin-BSA conjugate in PBS for 2 hours at 37°C, then saturated with 2% BSA in PBS for 1 hour at 37°C, and washed 3x with 0.05% Tween-PBS. The hybridoma supernatants to be screened are diluted in a 1% solution of BSA in PBS, and incubated overnight at room temperature (or 18 hours at 4°C or 2 hours at 37°C). Level of bound antibody is measured by anti-mouse IgG goat globulin coupled to alkaline phosphatase with para-nitrophenylphosphate as the substrate. After incubation for two hours at 37°C, the enzymatic substrate is hydrolysed (i hour at room temperature) and absorbance at 405 nm is measured. Hybridomae are selected for production of high affinity monoclonal antibody.

Standard curves to determine relative affinity of a selected antibody to rapamycin are prepared using solutions containing known concentrations of rapamycin (e.g., 1 to 140

ng/ml in blood serum). Figure 2, for example, shows a standard curve for our monoclonal antibody M7-91, demonstrating that that monoclonal antibody, which was selected as being highly specific for rapamycin, is capable of detecting rapamycin at levels as low as 0.25 ng/ml.

Antibodies may be further characterized as binding to the effector or FKBP binding domains of rapamycin by measuring cross reactivity with FK-506 in an analogous direct ELISA using microtiter plates coated with FK506-BSA conjugate, which can be prepared analogously to rapamycin-BSA conjugate. For example, a comparison of binding levels of 17 selected monoclonal antibodies in the rapamycin-BSA and FK-506 assays is shown in figure 3; cross reactivity as a percentage is shown in figure 4. In this comparison of binding to rapamycin-BSA vs. FK506-BSA, monoclonal antibodies of very low affinity are detected.

The above direct ELISA may be converted to a competitive ELISA wherein a competitor is added to the monoclonal antibody solution, and binding of the monoclonal antibody to the conjugate in the presence and absence of the competitor is measured. Where the competitor is FK506 or a rapamycin, the competitor in ethanolic solution, e.g., 1 mg/ml is directly added to the monoclonal antibody solution (e.g., 2 μ l/200 μ l/well) and further diluted in the microtiter plate. Figure 5, for example, shows an inhibition curve for binding of the M7-91 antibody (M7.91.13) to BSA-rapamycin in the presence of different concentrations of free rapamycin. In such competitive ELISAs comparing the binding of the monoclonal antibodies to free FK506 vs. free rapamycin, less cross reactivity between rapamycin and FK506 is seen than in the direct ELISA. Such a competitive assay is preferred for selection of monoclonal antibodies, because it is believed that some monoclonal antibodies may have too low an affinity to bind their antigen in free form in solution but may nevertheless show bivalent or polyvalent binding to the multimeric antigen, which comprises many haptens bound in close proximity to one another on a large protein molecule. A competitive assay excludes such low affinity antibodies. The results of such a competitive assay are shown in figure 6, which compares the M7-91 antibody (M7.91.13) which has relatively low cross reactivity with M1-303 (M1.303.3)

antibody which has relatively high cross reactivity.

b) *ELISA for 40-O-(2-hydroxyethyl)-rapamycin*

This ELISA is performed analogously to the procedure described in a). Microtiter plates are coated with 40-O-(2-hydroxyethyl)-rapamycin-BSA, saturated with BSA and washed. The hybridoma supernatants to be screened are incubated 18 hours at 4°C or 2 hours at 37°C. Level of bound antibody is measured by anti-mouse IgG goat globulin coupled to alkaline phosphatase with para-nitrophenylphosphate as the substrate. A parallel ELISA is performed using bound rapamycin-BSA to select for monoclonal antibodies capable of distinguishing 40-O-(2-hydroxyethyl)-rapamycin from rapamycin. Figure 7 shows, for example, supernatants of hybridoma B3-203 binding selectively to 40-O-(2-hydroxyethyl)-rapamycin-BSA (referred to in the figure as BSA-28-RAD) (figure 7A), of hybridoma B3-113 recognizing both 40-O-(2-hydroxyethyl)-rapamycin-BSA and rapamycin-BSA conjugated through position 28 (figure 7B), and of hybridoma B3-164 that recognize in addition rapamycin coupled to BSA through position 40 (figure 7C).

The relative affinity of the antibodies to 40-O-(2-hydroxyethyl)-rapamycin versus rapamycin is further measured by competing the binding of the antibodies to the coated 40-O-(2-hydroxyethyl)-rapamycin-BSA conjugate with 40-O-(2-hydroxyethyl)-rapamycin or rapamycin free in solution. Figure 8, for example, shows that antibodies produced by hybridoma B3-203 react strongly with 40-O-(2-hydroxyethyl)-rapamycin with low crossreactivity for rapamycin (figure 8A) and that antibodies produced by hybridoma B3-113 and B3-164 bind equally well to 40-O-(2-hydroxyethyl)-rapamycin and rapamycin (figures 8B and 8C). Other hybridomae producing antibodies binding at least 10-100-fold better to 40-O-(2-hydroxyethyl)-rapamycin than to rapamycin include B3-22, B3-127 and B3-156. Other hybridomae such as B3-29, B3-265 and B3-539 produce antibodies that bind rapamycin as well as 40-O-(2-hydroxyethyl)-rapamycin.

Once the desired antibody has been selected, the same ELISA is used to determine blood levels of rapamycin in patients. An assay kit according to this example would provide one or more selected antibodies in lyophilized form, a microtiter plate coated with

a rapamycin conjugate (e.g., rapamycin-BSA conjugate or 40-O-(2-hydroxyethyl)-rapamycin-BSA conjugate), a rapamycin standard, and instructions for use. Optionally, the kit may further comprise a labelled rapamycin derivative for use in a competitive assay. Anti-mouse IgG-enzyme conjugate and substrate as described above may additionally be provided. Alternatively, the customer may use the monoclonal antibody of the invention in his own established ELISA or other assay system.

CLAIMS

1. A monoclonal antibody capable of specifically recognizing a rapamycin.
2. A monoclonal antibody according to claim 1 capable of recognizing an epitope on the FKBP-binding portion of a rapamycin.
3. A monoclonal antibody according to claim 1 capable of recognizing an epitope on the effector portion of a rapamycin.
4. A monoclonal antibody according to any one of claims 1 through 3, obtained or obtainable by:
 - a) reaction of a rapamycin having an activated coupling group with an immunogenic protein to produce an immunogenic conjugate,
 - b) administration of said immunogenic conjugate to an appropriate animal species to effect immunogenic challenge and recovery of antibody-producing cells sensitized to said conjugate,
 - c) immortalization of said antibody-producing cells, and
 - d) recovery of monoclonal antibody from a selected immortalized cell line thus established.
5. A monoclonal antibody according to any one of claims 1 through 4 wherein the rapamycin is (i) rapamycin; or (ii) a 40-O-alkylated rapamycin, preferably wherein the 40-O-alkyl substituent is selected from hydroxyalkyl, hydroxyalkoxyalkyl, acylaminoalkyl, or aminoalkyl.
6. A monoclonal antibody according to claim 5 wherein the rapamycin is selected from
 - i) 40-O-(2-hydroxyethyl)-rapamycin,
 - ii) 40-O-(3-hydroxypropyl)-rapamycin,

- iii) 40-O-[2-(2-hydroxy)ethoxy]ethyl-rapamycin, and
- iv) 40-O-(2-acetaminoethyl)-rapamycin).

7. A monoclonal antibody according to any of the preceding claims capable of distinguishing between rapamycin and a 40-O-alkylated rapamycin as described in claim 5 or 6.

8. An immunogenic conjugate comprising a rapamycin portion and a protein portion.

9. An immunogenic conjugate according to claim 8 which is produced by reacting a protein with a rapamycin derivative bearing an activated coupling group.

10. An immunogenic conjugate according to claim 9 wherein the rapamycin derivative is selected from

- i) 40-O-succinimidooxysuccinyl-rapamycin,
- ii) 28-O-succinimidooxysuccinyl-rapamycin, or
- iii) 28-O-(succinimidooxysuccinyl)-40-O-(2-hydroxyethyl)-rapamycin.

11. A rapamycin having an activated coupling group.

12. A rapamycin according to claim 11 selected from

- i) 40-O-succinimidooxysuccinyl-rapamycin,
- ii) 28-O-succinimidooxysuccinyl-rapamycin, or
- iii) 28-O-(succinimidooxysuccinyl)-40-O-(2-hydroxy)ethyl-rapamycin.

13. A hybridoma cell line capable of producing a monoclonal antibody as defined in any one of claims 1 through 7.

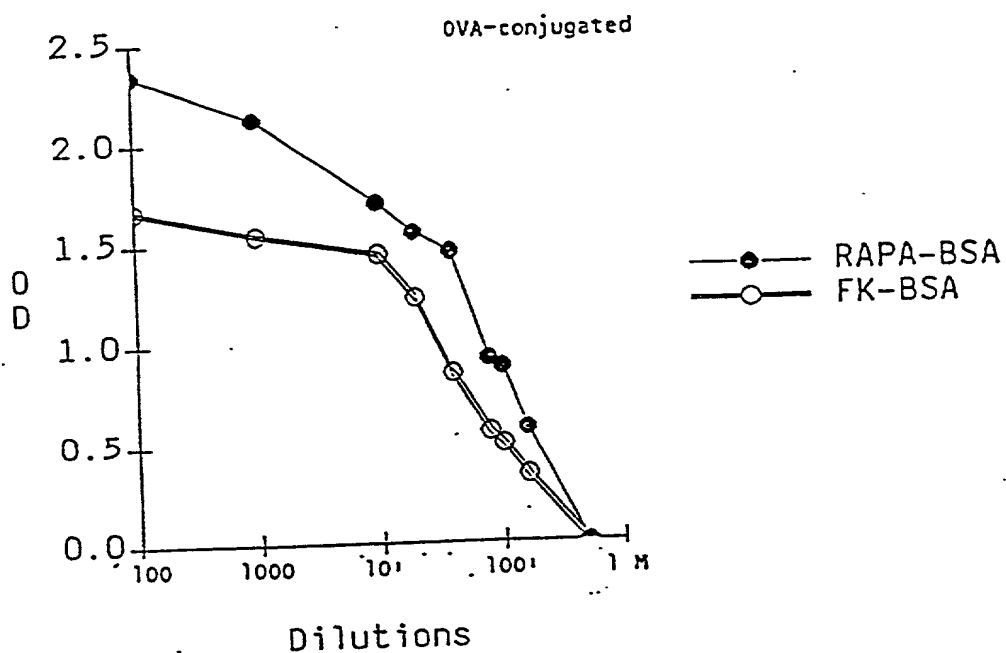
14. An immunoassay kit for measuring the blood level of a rapamycin, comprising a monoclonal antibody as defined in any one of claims 1 through 7.

ABSTRACT

Monoclonal antibodies to rapamycin and to 40-O-alkylated derivatives of rapamycin are provided, together with novel haptens, immunogenic conjugates, and processes for making them and assay kits for using them.

002227-2425550

Titer mouse M1 (RAPA/FK)



Titer mouse M7 (RAPA/FK)

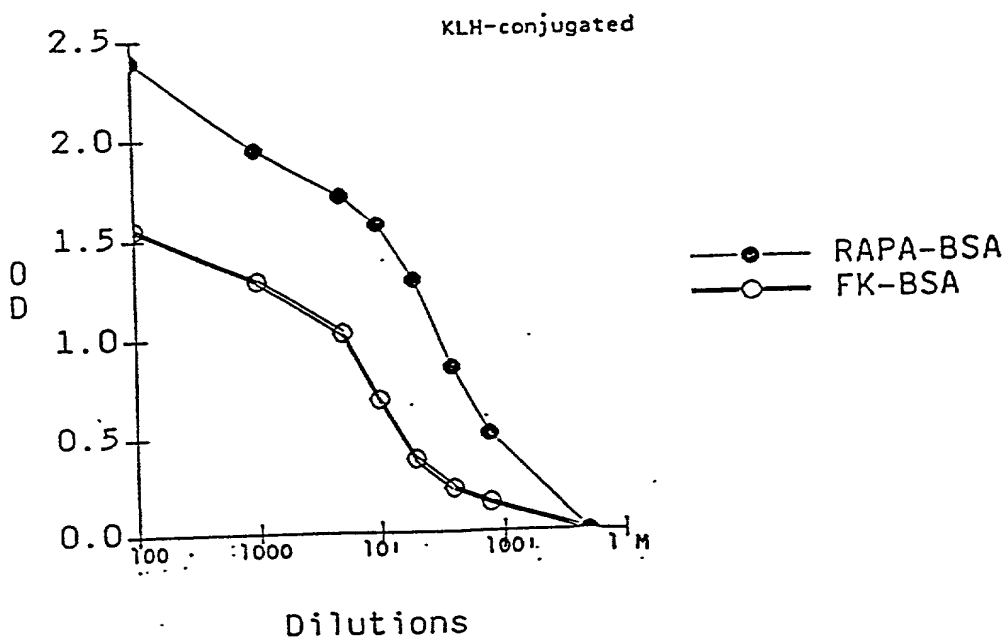


FIGURE 1

00000-ETH-5500

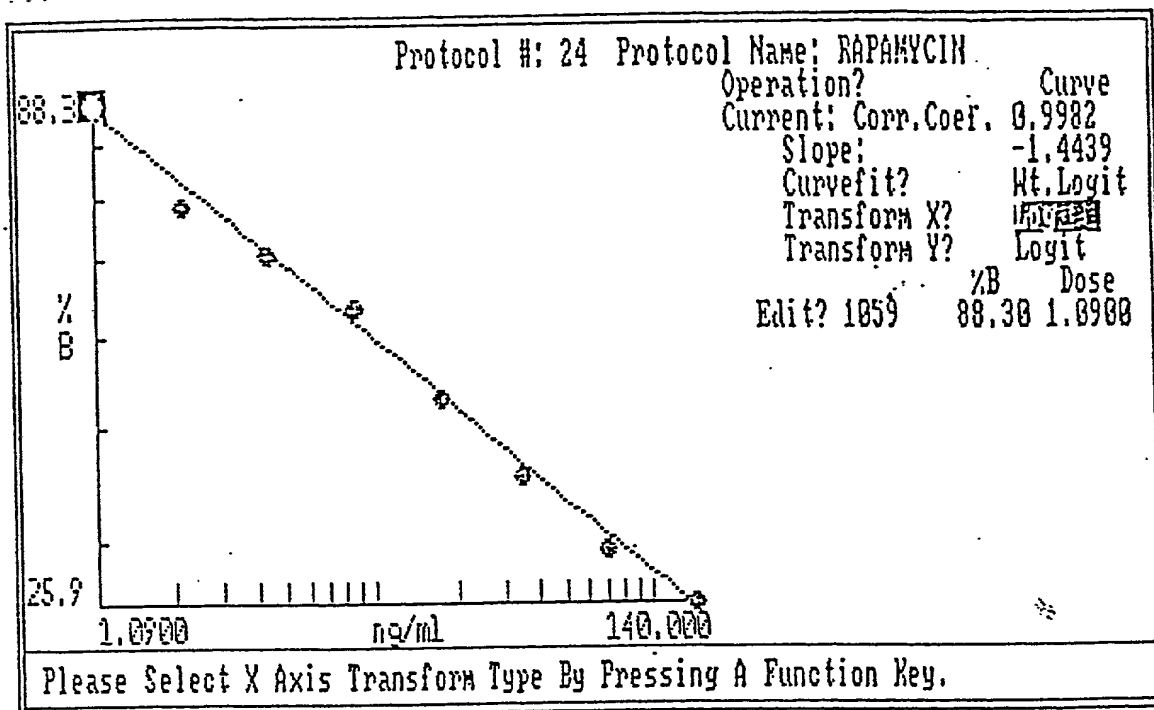


FIGURE 2

Screening of RAPA monoclonal antibodies RAPA/FK 506 Elisa

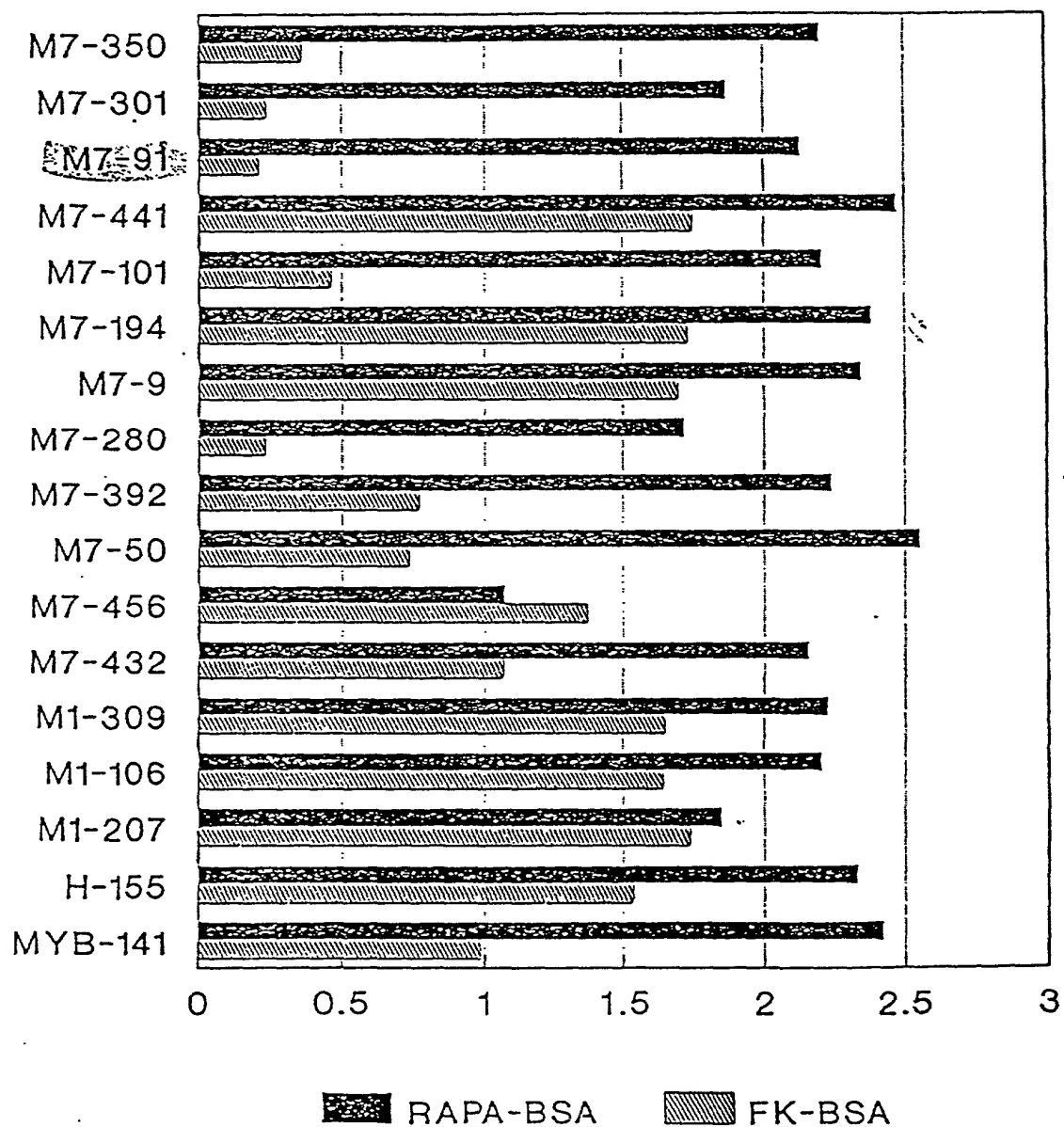


FIGURE 3

Screening of RAPA monoclonal antibodies

RAPA/FK 506 Elisa

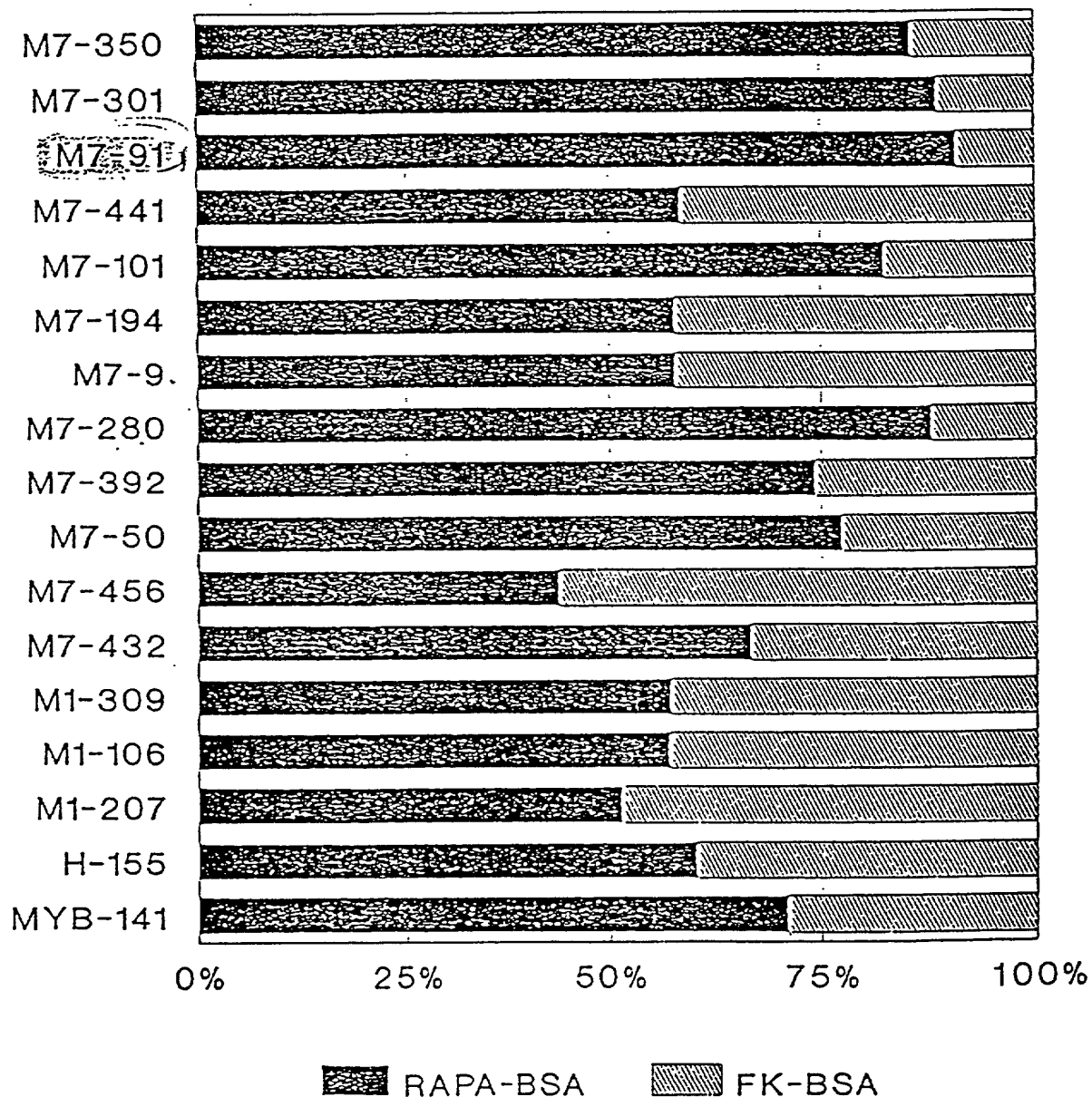


FIGURE 4

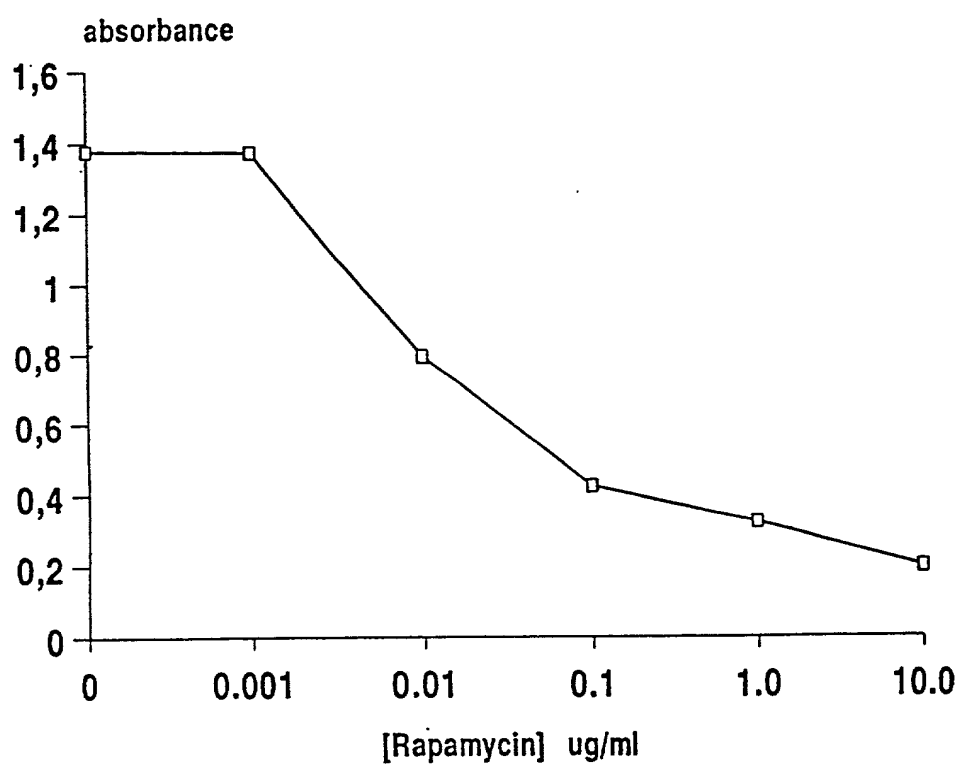
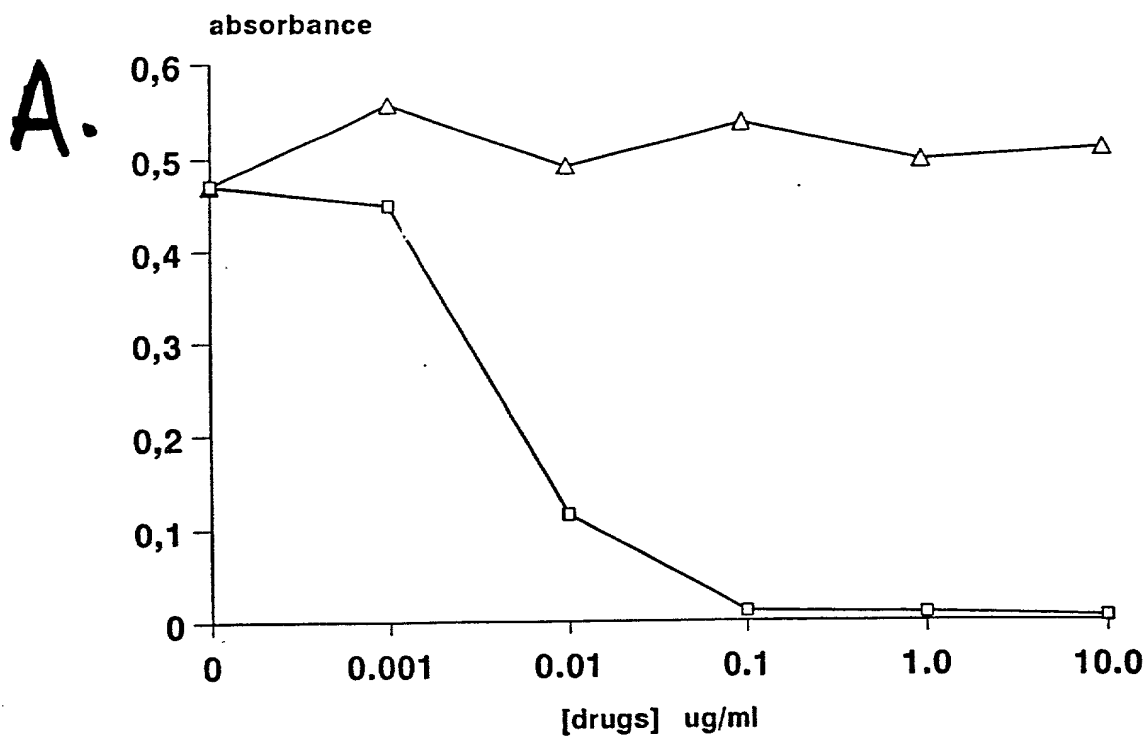


FIGURE 5

MAb M7.91.13



MAb M1.303.3

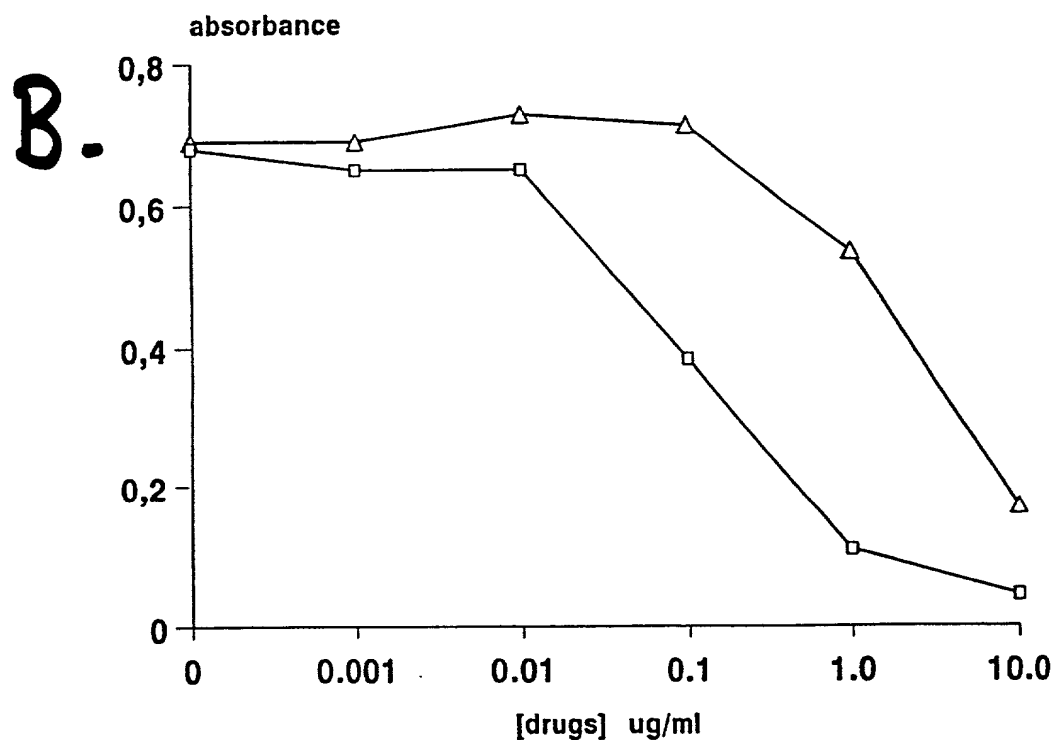
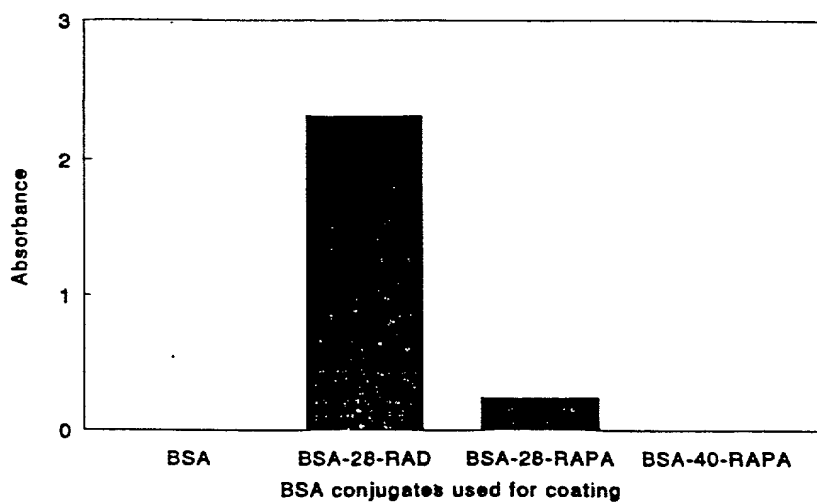
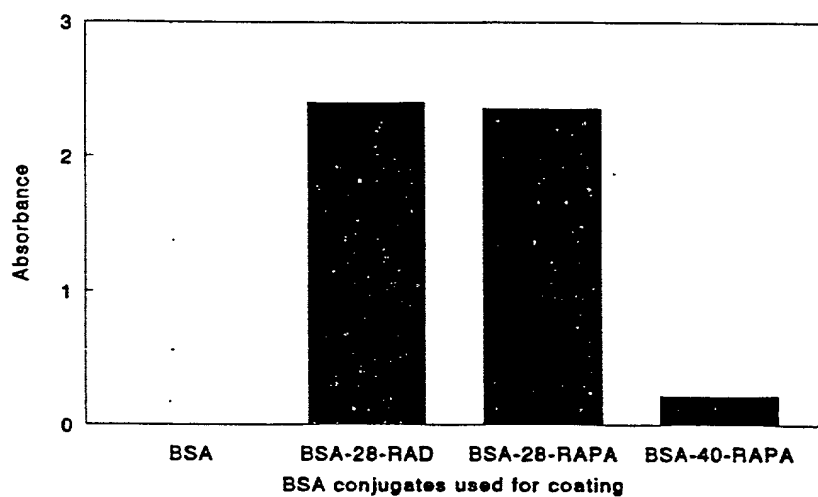


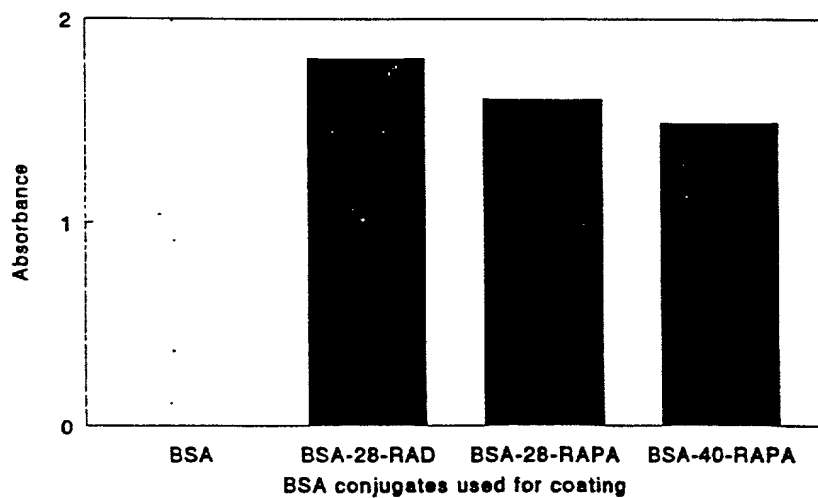
FIGURE 6 : Specificity of mAbs to Rapamycin for Rapamycin (□) vs. FK 506 (Δ) in competitive ELISA.



A

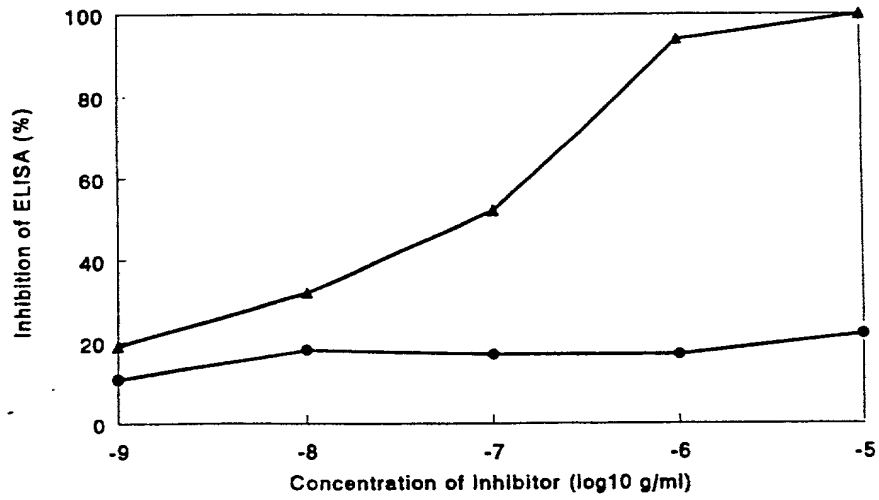


B

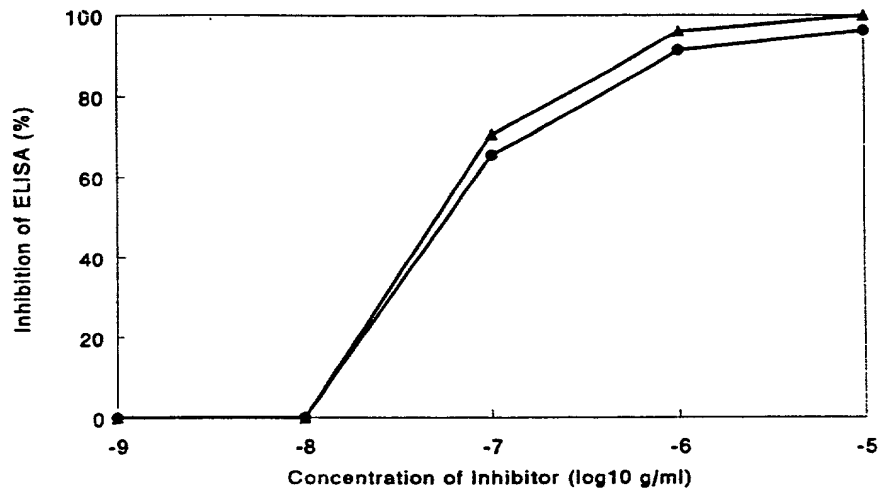


C

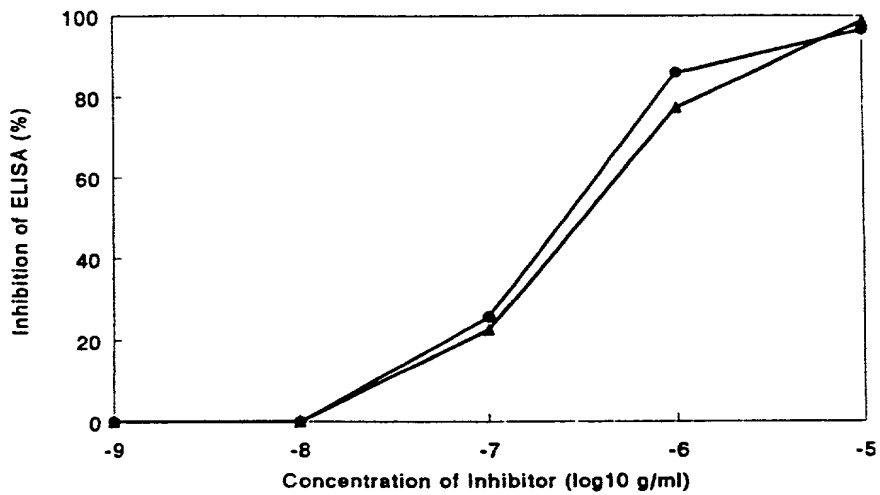
FIGURE 7



A



B



C

FIGURE 8

DECLARATION AND POWER OF ATTORNEY FOR
UNITED STATES PATENT APPLICATION

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name, and

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if more than one name is listed below) of the subject matter which is claimed and for which a United States patent is sought on the invention entitled

RAPAMYCIN ASSAY
the specification of which

☐ is attached hereto.

☐ was filed on , 19 as application Serial No. and, if these brackets contain an X ☐, was amended on ,

☒ was filed as Patent Cooperation Treaty international application No.

PCT/EP94/01006 on March 30, 1994, if these brackets contain an X ☐,

was amended under Patent Cooperation Treaty Article 19 on

, if these brackets contain an X ☐, entered the national stage in the

United States on , 19 and was accorded Serial No. /

, and, if these brackets contain an X ☐, was amended on

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment(s) referred to above.

I acknowledge my duty to disclose all information which is known by me to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I hereby claim the benefit under Title 35, United States Code, §119(a)-(d) of any foreign application(s) for patent or inventor's certificate indicated below and of any Patent Cooperation Treaty international application(s) designating at least one country other than the United States indicated below and have also identified below any foreign application(s) for patent or inventor's certificate and Patent Cooperation Treaty international application(s) designating at least one country other than the United States for the same subject matter and having a filing date before that of the application for said subject matter the priority of which is claimed:

<u>Country</u>	<u>Number</u>	<u>Filing Date</u>	<u>Priority Claimed</u>
Great Britain	94307491.2	April 8, 1993	[X] Yes [] No
			[] Yes [] No
			[] Yes [] No
			[] Yes [] No
			[] Yes [] No

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below:

Application

<u>Serial No.</u>	<u>Filed</u>

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and of any Patent Cooperation Treaty international application(s) designating the United States listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in said prior application(s) in the manner required by the first paragraph of Title 35, United States Code, §112, I acknowledge my duty to disclose all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date(s) of the prior application(s) and the national or Patent Cooperation Treaty international filing date of this application:

<u>Application</u>	<u>Status (Pending, Abandoned or Patented)</u>
<u>Serial No.</u>	<u>Filed</u>

I hereby appoint the following:

ROBERT S. HONOR	Reg. No. 22,801
THOMAS O. MCGOVERN	Reg. No. 25,741
MELVYN M. KASSENOFF	Reg. No. 26,389
JOSEPH J. BOROVIAN	Reg. No. 26,631
DIANE E. FURMAN	Reg. No. 31,104
CARL W. BATTLE	Reg. No. 30,731
JOHN L. CHIATALAS	Reg. No. 31,818
CAROL A. LOESCHORN	Reg. No. 35,590
MICHAEL P. MORRIS	Reg. No. 34,513
THOMAS C. DOYLE	Reg. No. 22,340

respectively and individually, as my attorneys and/or agents, with full power of substitution and revocation, to prosecute this application and to transact all business in the Patent and Trade-mark Office connected therewith. Please address all communications to ROBERT S. HONOR, SANDOZ CORPORATION, 59 Route 10, East Hanover, New Jersey 07936-1080, whose telephone number is 201-503-8485.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Sole inventor or

first joint inventor: Full name : 3 August 1935
 Signature : Richard Sedrani
 Date : Richard Sedrani
 Citizenship : Luxembourg
 Residence : Herrengrabenweg 15, CH-4054 Basle,
 Switzerland
 P.O. Address : same as above

IMPORTANT: Before this declaration is signed, the patent application (the specification, the claims and this declaration) must be read and understood by each person signing it, and no changes may be made in the application after this declaration has been signed.

Second joint inventor,

if any:

Full name : 9. August 95
 Signature : V. Quesniaux
 Date : Valerie Quesniaux
 Citizenship : France
 Residence : Schützweg 7, CH-4051 Basle, Switzerland

P.O. Address : same as above

Third joint inventor,

if any:

Full name :
 Signature : _____
 Date : -
 Citizenship :
 Residence :

P.O. Address :

Fourth joint inventor,

if any:

Full name :
 Signature : _____
 Date :
 Citizenship :
 Residence :

P.O. Address :

Fifth joint inventor,

if any:

Full name :
 Signature : _____
 Date :
 Citizenship :
 Residence :

P.O. Address :